Diagnostic efficacy of the new prospective biomarker's combination CA 15-3 and CA-62 for early-stage breast cancer detection: Results of the blind prospective-retrospective clinical study

Janneta Tcherkassova^{a,∗}, Anna Prostyakova^b, Sergey Tsurkan^a, Vladislav Ragoulin^a, Alexander Boroda^c and Marina Sekacheva^c

a *JVS Diagnostics LLC, Skolkovo IC, Moscow, Russia*

^b*Shemaykin-Ovchinnikov Institute of Bioorganic Chemistry RAS, Moscow, Russia*

^c*World-Class Research Center "Digital Biodesign and Personalized Healthcare", Sechenov First Moscow State Medical University, Moscow, Russia*

Abstract.

BACKGROUND: Combination of different cancer markers is often used for predicting tumor growth, for the response to cancer therapy, and for increase in the positive diagnosis ratio in the malignant tumors.

OBJECTIVE: Evaluation of the diagnostic efficacy of CA 15-3 and CA-62 cancer markers combination for early stages of breast cancer (BC) detection.

METHODS: This blind study was performed on 2 clinically validated Sets that included serum measurements of CA 15-3 ELISA and CLIA-CA-62 assays in 488 serum samples with TNM classification. A study included 300 BC patients (254 at Stages I and II, 20 with ductal carcinoma in situ (DCIS), and 26 Stages III and IV patients), 47 patients with breast benign diseases, and 141 healthy controls.

RESULTS: Sensitivity for DCIS & Stage I breast cancer detection was 75% at 100% Specificity (AUC = 0.895) using a following combination of two antigens: $10 <$ CA15-3 $<$ 46 U/ml and CA-62 \geq 6300 U/ml, which allows eliminating false positive results. CONCLUSIONS: The results obtained in a blind study demonstrate that a combination of CA15-3 with CA-62 yields 75% Sensitivity at 100% Specificity for DCIS and Stage I breast cancer detection, which has a potential to be integrated into existing screening programs. Prostyakova^b, Sergey Tsurkan^a, Vladisla
Sekacheva^c
C, *Moscow, Russia*
of *Bioorganic Chemistry RAS, Moscow, Russigital Biodesign and Personalized Healthcare*
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Keywords: Breast cancer (BC), Stage I, CA 15-3, CA-62, cancer marker

1. Introduction

Global cancer statistics confirms that in the last decade female breast cancer has become the most prevalent cancer in women, with more than 2 million cases worldwide corresponding to 11.7 cancer incidence [\[9\]](#page-12-0). At the same time the global epidemiological reports [\[20\]](#page-12-1) indicate that the breast cancer mortality rate has increased significantly in the last decade demonstrating an annual overall increase by 0.23%. Majority of the global breast cancer patients are diagnosed in the USA, Canada, Mexico, South America, Nordic countries, France, Switzerland, Israel, and the Netherlands [\[26\]](#page-12-2). In Russia, around 50,000 new BC cases are diagnosed every year, and approximately 30% of them

[∗]Corresponding author: Janneta Tcherkassova, JVS Diagnostics, Skolkovo, Bolshoi Boulevard 42/1, Moscow, Russia. Tel.: +7 9663188683; E-mail: janneta_tcherkassova@oncomarker.ru.

are detected at advanced stages of cancer, which causes high net mortality rate. Cumulative risk of the breast cancer development is equal to 5.4% for a woman's lifetime, and the incidence rate of breast cancer increases with age from less than 0.5% at 40 years of age to 2.86% at 60 years of age. Some other factors, such as hereditary breast and ovarian syndrome caused by BRCA1 & BRCA2 mutations, age-related changes in the female sex hormones levels up to 10 times increase the risk of breast cancer in this risk group than the baseline [\[9](#page-12-0)[,25\]](#page-12-3).

Women with breast cancer diagnosed at Stage I have 98 percent or higher survival rate in the first five years [\[2,](#page-12-4)[9\]](#page-12-0). Obviously that non-invasive ductal carcinoma in situ (Stage 0) and early stages of breast cancer have better outcomes than more advanced stages of cancer. Over the course of the last two decades, a number of achievements have taken place in breast cancer research that significantly improved the chances for patient's survival. Some significant advances in breast cancer screening methods, breakthroughs in cancer treatments, most recent achievements in molecular biology and molecularly targeted therapies provided better understanding of breast cancer development, enabling the design of the novel highly effective and safe treatment strategies. Breast cancer screening techniques include various tests such as mammography, magnetic resonance imaging, and ultrasound with or without elastography [\[2\]](#page-12-4). Currently there are a few ongoing clinical observational or prevention clinical trials for breast cancer such as BRCA-P (BCT 1801/ABCSG 50) or BCT 2001 (Breast MRI Evaluation) aimed to create the new methods to detect, precisely diagnose breast cancer, and reduce the risk of the disease for patients today and in the future. The extensive diversion of the search and the search and

The development of screening techniques for early detection of breast cancer is challenging due to high prevalence of the disease and the differentiation of benign and malignant breast tumors. The biggest drawback of the overall screening is related to the very high cost of the screening programs, which have significant budget implications, depending on the size of the population and the healthcare system resources involved [\[22\]](#page-12-5). Currently the most commonly used screening test for breast cancer in women without any symptoms is mammography, however it allows detecting only some fraction (from 20 to 60%) of Stage I breast cancer overlooking a majority of cases due to calcifications, imprecise aligning, biases, parenchyma density, and misinterpretations [\[4,](#page-12-6)[13](#page-12-7)[,21,](#page-12-8)[28\]](#page-12-9). As it turns out, a successful combination of various imaging methods such as mammography & MRI allows providing better detection of early stages of breast cancer (BC) as compared to single method's use.

Besides imaging methods of diagnostics developed over the past few decades, some immunoassays based on blood serum biomarkers provide a non-invasive promising approach for improving detection and monitoring of breast cancer [\[30\]](#page-12-10). For instance, a well-known cancer marker CA 15-3, an O-glycoprotein of the mucin family is commonly used for breast cancer control management [\[2](#page-12-4)[,28\]](#page-12-9). Despite its low sensitivity for early stages of breast cancer detection, CA 15-3 cancer antigen is extensively used for cancer treatment monitoring in combination with clinical examination and various imaging methods and for early detection of cancer recurrence [1,6]. As opposite to the mucin CA 15-3, a marker for epithelial carcinomas CA-62 have demonstrated its diverse clinical potential for early detection of epithelial cancers of the breast, prostate, lung, and colon for screening, cancer treatment monitoring, and cancer recurrence detection [\[11\]](#page-12-13). Heavily glycosylated N-glycoprotein CA-62 is a mesenchymal marker that is expressed in large quantities from the onset of cancerogenesis, starting from the abnormal cells production by non-invasive carcinomas in situ, which is the unique quality of this biomarker.

In this paper, we present the results of the clinical blind study to discuss the combined use of two glycoproteins CA 15-3 and CA-62 for Stage I breast cancer detection using an elevated cutoff value to eliminate false positives among healthy women. Overall, we assessed a combined use of CA 15-3 and CA-62 cancer markers for Stage I breast cancer detection and DCIS, as well as sensitivity (Sen), specificity (Sp), positive predictive value (PPV), negative predictive value (NPV), area under the curve (AUC), and test accuracy for early stages of breast cancer detection.

2. Materials and methods

2.1. Serum samples

All cancer serum samples were taken before the treatment administration. Tumor staging was performed according to the tumor, lymph node, metastasis (TNM) classification by AJCC 8th Edition [\[14\]](#page-12-14). The TNM System for describing the anatomical extent of a neoplasm is based on 3 components: T – the size or direct extent of the primary tumor, N – the degree of cancer spread to regional lymph nodes, and M – the presence of distant metastases. Histopathological classification of the

All sets	Breast cancer $N = 300$	Breast benign $N=47$	Healthy $N = 141$	Total $N = 488$			
Set 1 (blind)	$N = 257$	$N=0$	$N=69$	$N = 326$			
Set 2 (blind)	$N=43$	$N=47$	$N=72$	$N = 162$			
Age, years							
Mean	64 years	46 years	55 years	59 years			
Median	66 years	46 years	54 years	61 years			
Range	$24 - 93$	$24 - 72$	$20 - 85$	$20 - 93$			
Age group, $n(\%)$							
$<$ 50 years	49 (16,3%)	31(66%)	46 (32,6%)	126 (25,8%)			
from 50 to 60 years	48 (16%)	$12(25,5\%)$	51 (36,2%)	111(22,7%)			
from 60 to 70 years	103(34,3%)	$3(6,4\%)$	34 $(24,1\%)$	140 (28,7%)			
from 70 to 80 years	81 (27%)	$1(2,1\%)$	$8(5,7\%)$	90 (18,4%)			
> 80 years	$19(6,3\%)$	$0(0\%)$	$2(1,4\%)$	21(4,3%)			
Race, n $(\%)$							
Caucasian	264 (88%)	47 (100%)	141 (100%)	452 (92,6%)			
Asian	36 (12%)	$0\left(0\%\right)$	$0(0\%)$	36 (7,4%)			
Clinical cancer stage, n (%)							
DCIS(0)	20(6,7%)	N/A	N/A	N/A			
Stage I	157 $(52,3\%)$						
Stage II	97 (32,3 %)						
Stage III	13 $(4,3\%)$						
Stage IV	13 $(4,3\%)$						
Benign breast diseases, n (%)							
Fibroadenoma	23 (49%)	N/A	N/A	N/A			
Fibroid-cystic disease	10(21%)						
Mastitis	4(8,5%)						
Papilloma of the milk gland	3(6%)						
Other (calcification, inflammation, keratosis, breast cyst,	6(12%)						
galactorrhea etc.)							
Region, n $(\%)$							
USA	243 (81%)	47 (100%)	72 (51.1%)	362 (74.2%)			
Russian federation	57 (19.0%)	$0(0\%)$	69 (48.9%)	126(25.8%)			
tumors was used as the gold standard in breast		Stage II ($N = 14$), and Stage IV ($N = 13$) breast					
r diagnosis. The entire set of serum samples, in-		cer patients was obtained from the Bio Specimen					
ig Set 1 and Set 2 was separated by centrifugation		Precision For Medicine, Inc (USA). Majority of c.					
g, 10 minutes) in BD SST tubes with silica clot		samples (98%) from Sets 1 and 2 were from pat					
tor and separating gel, heat inactivated at 56° C for		with early invasive stages of breast cancer (Sta					
and II; $N = 254$) and non-invasive ductal carcinor n using standard operating procedures for serum							
situ (DCIS; $N = 20$). Majority (81%) of breast ca tion [10,19] and stored at -86° C until used. The							
in age of healthy individuals was 56 years of age patients was identified by screening test and 19%							
e 44–85 years) and 66 years of age for breast can- identified by clinical presentation.							
tients (range 24–93).		Blood from healthy control subjects ($N = 6$)					

Table 1 Baseline characteristics of the patients

breast tumors was used as the gold standard in breast cancer diagnosis. The entire set of serum samples, including Set 1 and Set 2 was separated by centrifugation (1300 g, 10 minutes) in BD SST tubes with silica clot activator and separating gel, heat inactivated at 56◦C for 30 min using standard operating procedures for serum collection [\[10](#page-12-15)[,19\]](#page-12-16) and stored at −86◦C until used. The median age of healthy individuals was 56 years of age (range 44–85 years) and 66 years of age for breast cancer patients (range 24–93).

Total of 488 blind serum samples from 300 histologically confirmed breast cancer patients with known TNM classification, 47 patients with benign breast diseases and 141 healthy individuals were included in this study. The study included 2 validated sets. Blind validated Set 1 included 326 serum samples including healthy controls ($N = 69$), Stage I ($N = 141$), Stage II ($N =$ 83), and Stage III ($N = 13$) breast cancer patients, as well as patients with ductal carcinoma in situ (DCIS, $N = 20$). Another blind validation set 2 contained 162 archived specimens including healthy controls $(N =$ 72), breast benign patients ($N = 47$), Stage I ($N = 16$), Stage II ($N = 14$), and Stage IV ($N = 13$) breast cancer patients was obtained from the Bio Specimen bank Precision For Medicine, Inc (USA). Majority of cancer samples (98%) from Sets 1 and 2 were from patients with early invasive stages of breast cancer (Stages I and II; $N = 254$) and non-invasive ductal carcinoma in situ (DCIS; $N = 20$). Majority (81%) of breast cancer patients was identified by screening test and 19% was identified by clinical presentation.

Blood from healthy control subjects $(N = 69)$ of a Set 1 as well as part of pre-treatment breast cancer serum specimens ($N = 57$) were collected and processed according to the standard approved protocol at the Sechenov First Moscow State Medical University ("Sechenov University"), Moscow, Russia, for another prospective study, processed and stored at −86◦C until used. Another part of breast cancer serum samples $(N = 200)$ included in Set 1 and Set 2, as well as healthy controls ($N = 72$) and breast benign ($N = 47$) samples were obtained from the Bio Specimen bank Precision For Medicine, Inc (USA). The baseline characteristics of the patients are presented in Table [1.](#page-2-0)

2.2. Ethics approval and consent to participate

The study was approved by the Ethics Committee of the Sechenov First Moscow State Medical University. All patients were given an informed consent to participate in the study. In total 57 patients with histologicallyconfirmed BC and 69 healthy subjects from the prospective study were included in the analysis. Serum samples were collected at the Sechenov University Hospital after overnight fasting and delivered to the Clinical laboratory. The rest of serum samples were collected as consented donor samples from Federal licensed/registered facility following GMPs. The protocol for the study was approved by the local IRB. The informed consent for archived samples from the bio bank was not required by IRB.

2.3. Statistical analysis

The distribution of CA-62 and CA 15-3 levels in sera from healthy, breast benign, and breast cancer patients were tested for normality using the histograms. The determination coefficient (r) was used to evaluate the correlation between cancer antigen's CA-62 and CA 15-3 serum levels. Since CA-62 values differ by 1000 times from CA 15-3 values, the original measurements for each cancer antigen in different sample cohorts were log transformed (log10) before the analysis for obtaining the same equivalent scales. The ROC curves for the comparison and the distribution of both tumor markers CA-62 and CA 15-3 for different stages of breast cancer and non-cancer samples were plotted using multiple variables graph tool with default settings of the MedCalc statistical software (version 19.7.4, MedCalc Software Ltd, Belgium, EU). A combination of two antigens was presented both in graphical and statistical formats. To plot the ROC curve for the combination of CA-62 and CA 15-3 cancer markers using defined parameters the following technique was developed: a conversion of two independent parameters, such as CA-62 and CA 15-3 values to one arbitrary value corresponding to the principles-based developed testing approach. The method is described in detail in the Results. Fraction intensions and the topes of CA 15-3 circle RB. The informed

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Diagnostic characteristics of CA 15-3 ELISA, CLIA-CA-62 assay, and their combination were evaluated based on sensitivity and specificity, test accuracy, PPV, NPV, and the ROC-curves AUC for different IVD methods were compared. The level of significance was set at $p < 0.001$. To classify a specimen as positive or negative, were used recommended cutoff values for single cancer antigens and a new combination of cutoff values for each antigen was determined that resulted in 100% specificity.

2.4. The sandwich CA 15-3 ELISA immunoassay

CA 15-3 solid-phase one-step sandwich ELISA (Federal Service for Surveillance in Healthcare of Russia (FSSH) – approved *in vitro* diagnostic (IVD) medical device from Vector-BEST, Novosibirsk, Russia, Serial No # T-8472) is designed for the quantitative measurement of the cancer antigen CA 15-3 bound between two types of monoclonal antibodies specific to different epitopes of CA 15-3 cancer antigen in human serum. Capture CA 15-3 specific monoclonal antibodies (Mabs) were immobilized in the wells of the microplate provided in the kit. CA 15-3 calibration standards, serum samples. Positive control, and a working solution of the secondary anti-CA 15-3-HRP Mab are then added to the wells that react with the antibodies to form the immune complex of the HRP-enzyme with Mab-CA-15-3 that generates measurable optical signal to be recorded by ELISA colorimetric reader. A linear calibration curve CA 15-3 is plotted for the quantitative measurement of the analyte concentration in unknown specimens. The intensity of the optical density (O.D) is directly proportional to the concentration of CA 15-3 antigen present in the analyzed specimens. Time-to-result: 3.5 Hrs. Sensitivity of the assay: 0.5 U/ml, a diagnostic range: 10–250 U/ml, a detection method: colorimetric. The cut-off value (30 U/ml) was recommended by the manufacturer based on CA 15-3 levels in sera from healthy females ($N = 97$) of 18–50 years of age.

2.5. Human CLIA-CA-62 immunoassay

The Human CLIA-CA-62 assay (FSSH – approved IVD medical device from JVS Diagnostics LLC, Moscow, Russia, Serial No # CLIA-CA-62-200221) is designed for the quantitative measurement of the carcinoma-specific antigen CA-62 in human blood serum for the detection of various epithelial carcinomas in the adult population, regardless of their gender and race. A set of reagents is based on a competitive solid-phase chemiluminescent immunoassay, where the competition for active binding sites of specific anti-CA-62 Mab, captured on the wells of the provided microplate carried out between the cancer antigen present in the analyzed sample and the luminescent CA-62 conjugate. Serum samples, CA-62 standards, Positive control, and CA-62 conjugate solution are then added simultaneously to the wells that react with the

capture antibodies to form the {anti-CA-62-Mab-(CA-62-Acridinium ether)} complex which produces light emission to be recorded by flash chemiluminescence reader. The intensity of the luminescent light emission is inversely proportional to the concentration of the analyte in the sample. A marker for epithelial carcinomas CA-62 is N-glycoprotein that is significantly elevated in patient's serum with primary epithelial carcinomas such as breast, prostate, lung, ovaries, and colon, including those with stage I disease. A series of Standard calibrators CA-62 are tested simultaneously with the samples for plotting a Logit-Log calibration curve for the quantitative determination of the glycoprotein CA-62 in measurement units (U/ml) in unknown samples. CLIA-CA-62 designed as an assisting method of *in vitro* diagnostics that is intended to help a doctor with clinical decision-making. It is recommended by the manufacturer for early cancer detection in combination with clinical information and other diagnostic procedures. Measurements were made following manufacturer's instructions. Sensitivity of the assay: 35 U/ml; Measurement range: 50–10,000 U/ml. The total analysis time is 3.5–4 hours. The cut-off value (5000 U/ml) was recommended by the manufacturer based on CA-62 levels in sera from 513 healthy individuals of 18–72 years of age.

3. Results

3.1. Measurement of CA 15-3 and CA-62 serum levels in breast cancer, breast benign patients, and healthy controls

The entire set of serum samples was analyzed using *in vitro* diagnostic FSSH – approved IVD medical devices, such as sandwich CA 15-3 ELISA assay and a competitive chemiluminescent immunoassay CLIA-CA-62 for CA-62 glycoprotein concentration measurement. Table [2](#page-4-0) shows the descriptive statistics of CA 15-3 and CA-62 values in the serum of various stages of breast cancer, benign breast diseases, and healthy controls. CA 15-3 values correlated with the literature data [\[12](#page-12-17)[,14\]](#page-12-14) for the early and advanced stages of breast cancer. Measuring serum levels of cancer markers CA-62 and CA 15-3 in 488 serum samples (300 samples from patients with histopathological confirmation of breast cancer, 47 samples from breast benign patients and 141 samples from apparently healthy women) have established that CA-62 antigen was found to be elevated (12286 U/ml) in 279/300 (93%) breast cancer patient's samples (111/300 (37%) elevated serum level for CA 15-3 marker) as compared to its level in healthy women (2679 U/ml) and women with breast benign diseases (4465 U/ml). As opposed to CA 15-3, serum values for CA-62 glycoprotein have demonstrated the highest mean value for DCIS (Stage 0) (12223 U/ml) and Stage I (11856 U/ml) of breast cancer, that were decreasing with more advanced stages: Stage II (10092 U/ml), and Stages III and IV (9850 U/ml). Using recommended cutoff values for CA 15-3 ELISA and the CLIA-CA-62 assay the sensitivity was calculated for all stages of breast cancer, including Stage 0 (DCIS). The results obtained are presented in Table [3.](#page-6-0) It is worth mentioning that whereas the sensitivity of CA 15-3 is lower for early stages (20%–37%) as compared to advanced stages of breast cancer (46%), a sensitivity for the marker for epithelial carcinomas CA-62 is very high in all stages, with its maximum at pre-invasive DCIS $(Sen = 95\%)$ and Stage I $(Sen = 97\%)$ breast cancer that is decreasing with more advanced BC stages: Sen $= 88\%$ for Stage II, and Sen $= 85\%$ for stage III.

Despite the CA-62 marker's close mean and median values, the difference in the sensitivity of CA-62 for various stages is significant. The p-values for each stage demonstrate that a correlation was significant ($p <$ 0.001) at 95%-alpha level. However, it does not explain the difference in sensitivity for Stage I vs Stage II vs Stage III and IV. Two-tailed p-values for the sensitivity comparison of different stages of the CA-62 test are: $p = 0.0002$ (Stage II vs DCIS & Stage I), $p = 0.0138$ (Stage III and IV vs Stage I), $p = 0.8842$ (Stage III and IV vs Stage II). For CA 15-3 all the values are not significant ($p > 0.05$): $p = 0.1321$ (Stage II vs DCIS & Stage I), $p = 0.2087$ (Stage III and IV vs Stage I), $p = 0.7908$ (Stage III and IV vs Stage II). Therefore, for the assessment of the tests performance we used the Receiver operating characteristic (ROC) analysis (Fig. 1), which is widely accepted as the most complete way for quantification and reporting accuracy in twogroup classification. ROC-curve analysis allows achieving comprehensive means for comparing different tests by providing information about all possible pairs of achievable sensitivity and specificity values. Figure [1](#page-6-1) demonstrates the difference in the assay performance in Stages I, II, III and IV. by canonical or the given
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3.2. ROC curve analysis

The comparison of ROC curves for both tumor markers CA-62 and CA 15-3 for different stages of breast cancer and non-cancer samples (breast benign and

Table 3 Sensitivity of CA 15-3 ELISA and CLIA-CA-62 assays for different stages of breast cancer using recommended cutoff values Assay Cutoff Specificity Sensitivity

			All stages	DCIS	Stage I	Stage II	Stage III	Stage IV
CA 15-3 ELISA	30 Units/ml	95%	111/300 (37%)	$4/20(20\%)$	56/157 (36%)	40/97(41%)	6/13(46%)	5/13(38%)
AUC , <i>p</i> -value			0.720	0.720	0.722	0.812	0.843	0.862
			0.0002	0.0002	< 0.0001	< 0.0001	< 0.0001	< 0.0001
CLIA-CA-62	5000 Units/ml	95%	279/300 (93%)	19/20 (95%)	153/157 (97%)	85/97 (88%)	11/13(85%)	11/13 (85%)
AUC , <i>p</i> -value			0.976	0.976	0.992	0.944	0.925	0.967
			< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001

Fig. 1. Combined ROC curves of CA 15-3 (a) and CA-62 (b) cancer markers for each stage.

healthy control subjects) were plotted using graph tool with default settings of the MedCalc statistical software. To classify a sample as positive or negative, originally were used recommended cutoff values for single cancer antigens and later was determined a new combination of cutoff values for each antigen that resulted in 100% specificity. Figure [1](#page-6-1) shows the CA 15-3 (a) and CA-62 (b) ROC curves for Stage 0 (DCIS), Stage I, Stage II, and Stages III and IV. The overall performance of the CA 15-3 ELISA was lower $(AUC = 0.72)$ and rather expected for early stages of breast cancer (Sen $= 37\%$) and for more advanced stages (Sen $= 46\%$). The capacity to discriminate breast cancer from non-cancer controls was more noticeable for CLIA-CA-62 assay with $AUC = 0.96$, especially for non-invasive Stage 0 (DCIS) and Stage I ($AUC = 0.976$ for CA-62 vs AUC $= 0.759$ for CA 15-3) (Table [4\)](#page-7-0).

To plot the ROC curve for the combination of CA-62 and CA 15-3 cancer markers using defined parameters was implemented the following mathematical technique: a conversion of two independent parameters, such as CA-62 and CA 15-3 values, to one arbitrary value corresponding to the principles-based developed testing approach in accordance with the following mathematical transformations:

- (a) Samples with $10 <$ CA15-3 $<$ 30 were assigned the actual values of more sensitive CA-62;
- (b) Samples with CA $15-3 > 30$ U/ml were assigned the values equal to:

$$
\frac{\text{CA }15\text{-}3}{30} * 6300 \; [U/ml]
$$

(c) Samples with CA $15-3 < 10$ U/ml were assigned the values equal to:

$$
\frac{\text{CA }15\text{-}3}{10} * 6300 [U/ml]
$$

1. Standard Error, DeLong et al., 1988. 2. 95% Confidence Interval, Binomial exact.

Therefore, samples were considered as "positive" if: CA15-3 > 30 U/ml, $10 <$ CA15-3 < 30 U/ml and $CA-62 > 6300$ U/ml, and "negative" otherwise. This approach allows the samples to have only one parameter to be considered. The ROC curves were plotted in accordance with the TNM classification. Diagnostic characteristics of CA15-3 ELISA, CLIA-CA-62 assay, and a combination of CA15-3 & CA-62 tumor markers were evaluated based on their sensitivity and specificity, test accuracy, PPV, NPV, and (ROC) curves were compared for different IVD methods. The level of significance was set at $P < 0.001$. These data are presented in Table [5](#page-7-1) and Fig. [2.](#page-7-2) The ROC curve analysis confirms our hypothesis that a combination of biomarkers selected produces better results for DCIS & Stage I detection than CA 15-3 cancer marker alone (AUC 0.895 vs 0.752) and other previously proposed panel of markers containing PENK, pro-SP, hGH and CA15-3 ($AUC =$ 0.785) [\[13\]](#page-12-7). Sensitivity, % Special

118/300 (39.3%)

10 < CA 15-3 < 30 U/ml

239/300 (79.7%)

10 < CA 15-3 < 46 U/ml

226/300 (75.3%)

10

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3.3. Combination of serum values of CA 15-3 and CA-62 cancer antigens

Given the incidence rate of breast cancer and the limited sensitivity (60–90%) of the mammography commonly used for breast cancer detection caused by many possible influences, such screening approach produces a large number of "false positive" results for women with "suspicious mammograms", which are incorrectly categorized as positive for the presence of disease. Pearson correlation analysis demonstrated low correlation coefficient between CA 15-3 and CA-62 values for the entire set of samples ($r = 0.2132$, $p < 0.001$), which

Fig. 2. ROC-curve for Stage I with DCIS using a combination of CA 15-3 and CA-62.

indicates an absence of the linear regression between two variables. However, plotted correlation between CA 15-3 and CA-62 serum values revealed that the majority of cancer samples are "scattered" in one quadrant, located above 10 U/ml of the mucin CA 15-3 and above 6300 U/ml of CA-62 glycoprotein. That allowed us to set up a hypothesis suggesting that a combination of the two biomarkers selected could potentially improve early-stage breast cancer detection by considerable increase in sensitivity at 100% specificity. To maximally increase the detection of Stage I breast cancer, we have decreased a cutoff value for CA 15-3 cancer antigen to 10 U/ml and at the same time increased an established CA-62 cutoff value to 6300 U/ml in order to reach 100% Specificity that allows eliminating false positives results. As expected, such high specificity cutoff value reduces the sensitivity of CA-62 cancer marker alone, but at the same time brings the overall Sensitivity of

Fig. 3. A correlation between CA 15-3 and CA-62 cancer markers. A 100% Specificity cutoff value for CA 15-3 and CA-62 required a log scale on the x-values.

the biomarker's combination to clinically useful values $(Sen = 75\%)$.

We have determined the optimal combination for two biomarkers (Ca 15-3 $& CA-62$), which considers samples as "positive" if $10 < CA$ 15-3 < 30 U/ml and CA- $62 > 6300$ U/ml or CA 15-3 > 30 U/ml, and "negative" otherwise with 79.7% sensitivity at 96.8% specificity. The combination of CA-62 with CA 15-3 $\{10 < CA$ 15-3 < 46 U/ml and CA-62 > 6300 U/ml or CA 15-3 > 46 U/ml} at a 100% Specificity increased the sensitivity of the DCIS, Stage I and II breast cancer detection from 39% to 75% that were "invisible" by CA 15-3 assay alone (Fig. [3,](#page-8-0) Table [5\)](#page-7-1). Table [5](#page-7-1) demonstrates the diagnostic characteristics of the biomarkers selected for the entire studied set of breast cancer samples. Increasing the specificity of the biomarker's combination to 100% allows eliminating the false positive results occurrence, which usually harms screening programs. Alternative Section (and Research COPY)

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4. Discussion

In this paper we present the results of the blind study to discuss a combined use of two glycoproteins CA 15-3 and CA-62 for Stage I breast cancer detection using a very high cutoff value for both markers to eliminate false positives among healthy women. A combination of the IVD chemiluminescent CLIA-CA-62 and colorimetric CA 15-3 ELISA assays appears to offer prospective cost-effective solution for novel breast cancer primary screening strategy prior mammography.

Breast cancer is a silent killer, with very minor symptoms that are associated with yet invasive late-stages of the disease. The incidence rate of breast cancer is around 90 per 100,000 women in Western Europe and North America and around 57 per 100,000 women in Eastern Europe. With such frequency a successful screening strategy must provide a sensitivity of at least 75% at close to a 100% specificity to significantly decrease a probability of detecting false positive results. A number of screening approaches using cancer biomarkers and their panels were used in the past such as a screening study in the UK Collaborative Trial [\[15,](#page-12-18)[24\]](#page-12-19).

Currently the mammographic screening is the most common approach to breast cancer detection worldwide and it is the only one test clinically proven to decrease mortality rates [2]. Besides the obvious advantage of the mammography which allows detecting the malignant tumors less than 5 mm (26.9 % of BC cases) including some carcinomas in situ (DCIS), it has some significant drawbacks. For example, the mammographic screening of women is producing a significant amount (3/4) of "suspicious mammograms" (2.2%; 2200 per 100,000 women) that are not associated with breast cancer and considered as false positive results, decreasing a diagnostics efficacy of this screening strategy [\[20\]](#page-12-1). As it was demonstrated by authors [\[4](#page-12-6)[,23\]](#page-12-20) based on the results of the multicenter clinical trials, about 40% of BC cases are "missed" early-stage as a result of the detection and misinterpretation errors (dense parenchyma, calcifications, distortions, poor aligning etc). Therefore, the mammography detects only 63–90% of breast cancers in patients at screening that gives a probability of 10 to 37% that the breast cancer will not be detected. According to the WHO screening recommendations [\[5\]](#page-12-21) the overpriced economic costs to the health care system in regards to screening programs are related to a large number of false positive results that causes overdiagnosis, over-treatment, false positives, and strain on health systems. The quality of the screening test is described in terms of Sensitivity and Specificity. It is desirable that the screening test has a high PPV. On another hand, high NPV, a posteriori probability given a negative test result, reflects, first of all, the probability for someone to truly be healthy, in case of a negative test result, but not a reduction in costs. This leads to late cancer detection, which can be offset by regular screening with high sensitivity and specificity.

Therefore, there is a need in a new cost-effective approach that will result in a high positive predictive value (PPV) and a minimum rate of false positive results. It could be an improvement of the current mammographic approach using an additional instrumental method for visual detection of the vascularization associated with malignant tumors or a combination of the mammography with sensitive *in-vitro* diagnostic methods.

The results reported in this paper strongly suggest that the CLIA-CA-62 chemiluminescent assay is able to detect a significant increase in the CA-62 serum level of breast cancer patients in the DCIS, Stage I, Stage II, and Stages III and IV as compared to healthy women and women with benign breast lesions. The established overall sensitivity of CA 15-3 assay for breast cancer detection is in the range from 20 to 50%, which correlates with our findings (Sen $= 20-46\%$) with lower values for DCIS (20%) and Stage I (36%) and higher values for Stage II (41%) and Stages III and IV (46%). CA 15-3 has low sensitivity for detection the early stages of breast cancer, however it is widely used for cancer treatment monitoring of patients with advanced stages of breast cancer [\[16,](#page-12-22)18].

Unique quality of the marker for epithelial carcinomas CA-62 consists in its significant expression from the onset of cancerogenesis with its maximum production at non-invasive carcinoma in situ DCIS and Stage I breast cancer (Sen $= 95-97\%$), which decreases with the tumor progression: localized Stage II (Sen $= 88\%$) and regional or distant Stages III and IV (Sen $= 85\%$). Consider an absence of correlation between serum levels of CA 15-3 and CA-62 glycoproteins, a combination of two markers approach was proposed to determine whether it could potentially improve the sensitivity and the specificity of Stage I breast cancer detection. The results of the plotted values for CA 15-3 and CA-62

cancer markers demonstrated that the majority of cancer samples are "scattered" in one quadrant, located above 10 U/ml of the CA 15-3 marker and above 6300 U/ml for CA-62. Standard cut-off values for CA 15-3 ELISA $>$ 30 U/ml, and for CLIA CA-62 $>$ 5000 U/ml were used to determine the sensitivity and specificity, AUC area under curve, CI confidence interval for the individual assays. It is worth mentioning that the majority of the benign patients studied with CA-62 marker have demonstrated its baseline or slightly elevated level as compared to the high values for breast cancer specific cancer marker CA 15-3. Some of the breast benign specimens have demonstrated a slight increase in CA-62 level that might indicate a transitional stage of the tumor becoming malignant, which indeed was confirmed lately for some benign patients. The sensitivity, specificity, PPV, and accuracy of the CLIA-CA-62 IVD assay alone were 93%, 95%, 0,98, and 90%, which is approximately 1.5 times higher as compare to visual methods of diagnostics such as ultrasound, MRI, and mammography [2]. It is especially worth emphasizing the superior detection rate (97%) of the CLIA-CA-62 IVD assay from other cancer markers performance and their panels that are currently available for the detection of DCIS and Stage I of BC. Other methods of diagnostics for early-stage breast cancer have sensitivity from 27% for CA 15-3 assay alone [\[16](#page-12-22)[,18\]](#page-12-23) or 55% for multicancer early detection test MCED [\[6\]](#page-12-12) to 50% for mammography and 80% for MRI [\[8\]](#page-12-24). As it was demonstrated by [3] a screening of the pre-diagnosis 239 BC samples and cancer-free controls using 9 candidate biomarkers including CA 15-3, HSP90A and PAI-1, a combination models outperformed single markers, none of the candidates or their combinations were found useful for screening. Authors of another screening study [\[15\]](#page-12-18) have assessed the suitability of a biomarker panel (PENK, pro-SP, hGH and CA15-3) to improve early detection and individual risk assessment in breast cancer patients on 204 BC patients and 68 healthy controls. Their results for a biomarker panel were more promising: PENK and hGH concentrations were significantly lower and pro-SP values were higher in BC patients as compared with normal controls, whereas AUC increased from 0.628 for CA15-3 alone to 0.754 when was used a combination model of all three biomarkers. Frace of a negative

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> Comparison of different breast cancer diagnostic methods allows making a suggestion that only a combination of several methods is superior to the single use of either method for Stage I breast cancer detection providing a significant improvement in the DCIS and Stage I detection having high survival rate.

Our results validate that a combination of CA 15-3 with CA-62 biomarker could be potentially useful for precise differential diagnosis in breast cancer (DDx). A combination of two different N- and O-glycoproteins could significantly improve overall diagnostic results due to their specific features allowing successful differentiating benign from malignant breast tumors. Clinical value of CA 15-3 as a biomarker in breast cancer is very significant for management of BC patients, such as treatment monitoring and early detection of cancer recurrence [\[16\]](#page-12-22).

CA 15-3 is a secretory heavily O-glycosylated protein with high molecular weight that belongs to the MUCIN family, which secrets into the blood stream from the polysaccharide surface of the epithelial cells [\[16\]](#page-12-22). In cancer cells some structural changes of CA 15-3 occurred due to abnormal modifications in the glycosylation pattern to less glycosylated and more mobile form, inducing metabolic re-programming associated with tumor cell proliferative activity, resistance to hypoxia and stimulation of angiogenesis. Hyper expression of the mucin CA 15-3 occurs as the result of the Tumor necrosis factor (TNF) and Interleukin IL-8 production [\[25\]](#page-12-3). Increased expression of the mucin (CA 15-3) causes a suppression of the Tall-like receptors signaling and a decrease of the IL-8 expression. Therefore, understanding of the mucin expression role in cancer cells survival has an important impact on the development of the novel therapeutic, screening, and cancer treatment monitoring approaches for the successful treatment of breast cancer patients. A majority of other cancer markers including CA 15-3 represent heavy weight O-mucins (up to 800 kDa) that can get released into the blood only after tumor cells lysis. In this case, tumor cell death releases into the blood all the accumulated tumor-specific, tumor-associated and genetic markers which levels are directly proportional to the tumor growth. The that belongs to all ancinomas (substitute blues) to the bind the bind standard care and modifications in cosylated and more in CA-62 level than programming associativity, resistance of the tumor because in confirmed la

As opposed to mucins, a marker for epithelial carcinomas CA-62 belongs to a family of membrane Nglycoproteins that bind alpha-fetoprotein (AFP) using a special combination of oligosaccharides located on the surface of the antigen and function by receptormediated endocytosis [\[11\]](#page-12-13). N-glycoprotein CA-62 is a mesenchymal marker produced as a result of the malignant cells transformation by epithelial-mesenchymal transition (EMT) in which epithelial cells lose their characteristics (cell to cell junctions, their polarity, some epithelial markers) and acquire mobile cell shape and mesenchymal features. As a result, various epithelial cells possessing different morphological and phylogenetic classifications are transformed into the same pluripotent stem cells. Heavily branched Nglycoprotein is expressed in large quantities on the cell membrane of such degenerated mesenchymal stem cells from the onset of cancerogenesis, and is getting released into the blood stream from the cell membrane on the very early stages of cancer, including carcinomas in situ and detected by the CLIA-CA-62 chemiluminescent assay.

Previous studies have shown that malignant epithelial cells demonstrate significant expression of glycoprotein antigens CA-62 in all studied localizations of epithelial carcinomas (such as breast, prostate, lung, uterus, stomach, kidney, colon, and ovaries) [\[11\]](#page-12-13). At the same time, healthy controls do not demonstrate increased expression of CA-62 markers. However some breast benign specimens have demonstrated a slight increase in CA-62 level that might indicate a transitional stage of the tumor becoming malignant, which was actually confirmed lately for some benign patients. Patients with significant elevation in serum CA-62 level might have another type of carcinoma, which does not make it false positive for breast cancer detection, but rather beneficial for simultaneous detection of some other existing primary cancer.

Taken together the results of the biomarkers (CA 15- 3 and CA-62) combination may possibly be considered as indicator for early stages of BC and demonstrate their potential implementation into breast cancer screening programs. A proposed screening algorithm is presented in Fig. 4. Briefly, patients with $CA-62 > 5000$ U/ml will have to be monitored using a combination of cancer markers every 3 months within a year for the disease confirmation or its absence. Those patients with CA-62 level > 6300 U/ml and $10 <$ CA 15-3 $<$ 46 U/ml will be required to have additional instrumental visual diagnostic examination for the diagnosis confirmation. Therefore, a combination of CA 15-3 and CA-62 biomarkers could decrease an overall screening cost making such screening approach an economically viable solution.

Thus, the results obtained demonstrate that a combined use of serum CA 15-3 and CA-62 values provides high level of specificity (100%) and sensitivity (75%) suitable for the clinical detection of early stages of breast cancer, which could provide an effective tool for prior mammography breast cancer screening within generally healthy women above 35 years of age. This approach could also increase the diagnostic sensitivity in DCIS and Stage I breast cancer detection.

The indisputable advantage of using CA-62 cancer antigen for a biomarker's combination consists in its

Fig. 4. Proposed screening algorithm for target population over 30 years old using a combination CA-62 and CA 15-3.

very high sensitivity (Sen $= 97\%$) for early stages of cancer detection; however, it isn't breast cancer specific, which requires additional clarification. A combination of such sensitive carcinoma-specific antigen with breast cancer associated antigen, such as CA 15-3 could serve as a smart approach to screening for breast cancer.

In the context of this blind study, a biomarker's combination approach could prospectively be integrated into existing screening programs. The results obtained in the study confirm that by combining serum values of CA 15-3 and CA-62 it is possible to obtain a useful non-invasive screening test that will detect two-thirds of Stage I breast cancer without any false positives among healthy women. Such algorithm could supplement existing mammography screening as well as other diagnostic imaging methods. For the implementation of such biomarkers combination in the screening of BC our findings need to be confirmed in a clinical approbation.

5. Conclusions

The evidence from this study presents remarkable results in regards to the biomarker's combination for Stage 0 and Stage I breast cancer detection (Sen $=$ 75% at 100% Specificity), which is suitable for the clinical detection of early stages of breast cancer. The diagnostic efficacy of CA 15-3 cancer marker alone $(Sen = 20-39\%)$ can be significantly enhanced (75.3– 79.7%) by applying a second very sensitive biomarker CA-62 to a reduced 10 U/ml CA 15-3 cutoff value for the detection of DCIS, Stage I and II breast cancer at 95%–100% Specificity. A 100% Specificity value in the studied BC cohort samples was obtained at the following combination of two antigens: $10 < CA15$ - $3 < 46$ U/ml and CA-62 ≥ 6300 U/ml, which allows eliminating false positive results. An assessment of the results obtained using a combination of biomarkers (CA 15-3 and CA-62) demonstrate that such approach to BC detection has a potential in the nearest future to be integrated into existing screening programs.

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Author contributions

Conception: Tcherkassova J. Interpretation or analysis of data: Tcherkassova J, Ragoulin V. Preparation of the manuscript: Prostyakova A. Revision for important intellectual content: Tcherkassova J, Prostyakova A, Tsurkan S, Boroda A and Sekacheva M. Supervision: Tsurkan S, Sekacheva M.

References

- [1] A. Berruti, M. Tampellini, M. Torta, T. Bunieva, G. Gorzegno and L. Dogliotti, Prognostic value in predicting overall survival of two mucinous markers: CA 15-3 and CA 125 in breast cancer patients at first relapse of disease, *Eur J Cancer* 20(14) (1994), 2082–2084. doi: 10.1016/0959-8049(94)00356-A.
- [2] A. Bhushan, A. Gonsalves and J.U. Menon, Current state of breast cancer diagnosis, treatment, and theranostics, *Pharmaceutics* 13 (2021), 723–730. doi: 10.3390/pharmaceutics 13050723.
- [3] A. Kazarian, O. Blyuss, G. Metodieva et al., Testing breast cancer serum biomarkers for early detection and prognosis in pre-diagnosis samples, *Br J Cancer* 116 (2017), 501–508. doi: 10.1038/bjc.2016.433.
- [4] A. Majid, E. de Paredes, R. Doherty, N. Sharma and X. Salvador, Missed breast carcinoma: Pitfalls and pearls, *Radio-Graphics* 23(4) (2003), 881–895. doi: 10.1148/rg.234025083.
- [5] A short guide to cancer screening Increase effectiveness, maximize benefits and minimize harm, ISBN: 978-92-890-5756-1 c World Health Organization 2022. https://apps.who.int/iris/ bitstream/handle/10665/351396/9789289057561-eng.pdf.
- [6] B.W. Park, J.W. Oh, J.H. Kim, S.H. Park et al., Preoperative CA 15-3 and CEA serum levels as predictor for breast cancer outcomes, *Ann Oncol of Eur Soc Med Oncol (ESMO)* 19 (2008), 675–681. doi: 10.1093/annonc/mdm538.
- [7] E.A. Klein, D. Richards, A. Cohn et al., Clinical validation of a targeted methylation-based multi-cancer early detection test using an independent validation set, *Annals Oncology* 32(9) (2021), 1167–1177. doi: 10.1016/j.annonc.2021.05.806.
- [8] E.A. Morris, Rethinking breast cancer screening: Ultra FAST breast magnetic resonance imaging, *Journal of Clinical Oncology* 32(22) (2014), 2281–2283. doi: 10.1200/JCO.2014.56. 1514.
- [9] H. Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal and F. Bray, Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries, *Cancer J Clin* 71(3) (2020), 209–249. doi: 10.3322/caac.21660. 113-117, doi: 10

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- [10] J.F. Saluzzo, B. Leguenno and G. Van der Groen, Use of heat inactivated viral haemorrhagic fever antigens in serological assays, *J Virol Meth* 22 (1988), 165–172. doi: 10.1016/0166- 0934(88)90099-7.
- [11] J.R. Cherkasova, S.A. Tsurkan, V.B. Kondratiev and R. Moro-Vidal, Cancer antigen for early cancer detection, Patent WO2021/215955, 2021.
- [12] J. Wojtacki, A. Dziewulska-Bokiniec, J. Skokowski and D. Ciesielski, Evaluation of CA 15-3 tumor marker in the diagnosis of breast cancer. A pilot study, *Neoplasma* 41(4) (1994), 213–216. PMID: 7935994
- [13] K.K. Evans, R.L. Birdwell and J.M. Wolfe, If you don't find it often, you often don't find it: Why some cancers are missed in breast cancer screening, *PLoS One* 8 (2013), e64366.
- [14] M.B. Amin, AJCC cancer staging manual. Eight ed. 2017, ISBN 978-3-319-40617-6
- [15] M. Hamann, S. Grill, J. Struck et al., Detection of early breast cancer beyond mammographic screening: A promising biomarker panel, *Biomark Med* 13(13) (2019), 1107–1117. doi: 10.2217/bmm-2019-0085.
- [16] M.J. Duffy, Serum tumor markers in breast cancer: Are

they of clinical value? *Clin Chem* 52 (2006), 345–351. doi: 10.1373/clinchem.2005.059832.

- [17] M.J. Duffy, D. Evoy and E.W. McDermott, CA 15-3: Uses and limitation as a biomarker for breast cancer, *Clinica Chimica Acta* 411(23–24) (2010), 1869–1874. doi: 10.1016/ J.CCA.2010.08.039.
- [18] M.J. Duffy, S. Shering, F. Sherry, E. McDemmott and N. O'Higgins, CA 15-3: A prognostic marker in breast cancer, *Int J Biol Mark* 15 (2000), 330–333. doi: 10.1177/17246008000 1500410.
- [19] M.K. Tuck, D.W. Chan, D. Chia et al., Standard operating procedures for serum and plasma collection: Early detection research network consensus statement standard operating procedure integration working group, *J Proteome Res* 8(1) (2009), 113–117. doi: 10.1021/pr800545q.
- [20] N. Azamjah, Y. Soltan-Zadeh and F. Zayeri, Global trend of breast cancer mortality rate: A 25-year study, *Asia Pac J Cancer Prev* 20(7) (2019), 2015–2020. doi: 10.31557/APJCP. 2019.20.7.2015.
- [21] N. Cho, W. Han, B. Han et al., Breast cancer screening with mammography plus ultrasonography or magnetic resonance imaging in women 50 years or younger at diagnosis and treated with breast conservation therapy, *JAMA Oncol* 3(11) (2017), 1495–1502. doi: 10.1001/jamaoncol.2017.1256.
- [22] N. Mittmann, N.K. Stout, P. Lee et al., Total cost-effectiveness of mammography screening strategies, *Health Rep* 26(12) (2015), 16-25.7.
- [23] R.F. Brem, J. Baum, M. Lechner, S. Kaplan, S. Souders, L.G. Naul and J. Hoffmeister, American Journal of Roentgenology, 181 (2003), 687–693. doi: 10.2214/ajr.181.3.1810687.
- [24] S. Hofvind, B.M. Geller, J. Skelly and P.M. Vacek, Improvement in sensitivity of screening mammography with computeraided detection: A multiinstitutional trial, *The British Journal of Radiology* 85 (2012), e1226–e1232. doi: 10.1259/ bjr/15168178.
- [25] S. Joshi, S. Kumar, A. Choudhury, M.P. Ponnusamy and S.K. Batra, Altered Mucins (MUC) trafficking in benign and malignant conditions, *Oncotarget* 5(17) (2014), 7272–7284. doi: 10.18632/oncotarget.2370.
- [26] S. Lima, R. Kehm and M. Terry, Global breast cancer incidence and mortality trends by region, age-groups, and fertility patterns, *EClinicalMedicine* 38 (2021), 100985. doi: 10.1016/ j.eclinm.2021.100985.
- [27] S. McPhail, S. Johnson, D. Greenberg, M. Peake and B. Rous, Stage at diagnosis and early mortality from cancer in England, *Br J Cancer* 112 (2015), S108–S115. doi: 10.1038/bjc. 2015.49.
- [28] W. Zhao, X. Li, W. Wang et al., Association of Preoperative Serum Levels of CEA and CA15-3 with Molecular Subtypes of Breast Cancer, *Dis Markers* 2021 (2021), 5529106. doi: 10.1155/2021/5529106.
- [29] X. Castells et al., Risk of Breast Cancer in Women with False-Positive Results according to Mammographic Features, *Radiology* 280(2) (2016), 379–386. doi: 10.1148/radiol.2016151174.
- [30] X. Song, B. Liang, C. Wang and S. Shi, Clinical value of color Doppler ultrasound combined with serum CA153, CEA and TSGF detection in the diagnosis of breast cancer, *Experimental and Therapeutic Medicine* 20(2) (2020), 1822–1828. doi: 10.3892/etm.2020.8868.